

Compound 1 Colourless oil IR ν_{\max} cm^{-1} 3500, 1150, 1040; ^1H NMR (CDCl_3) δ 1.19 (3H, d, $J=6.7$ Hz), 1.26 (3H, s), 1.32 (3H, s), 1.49 (1H, dd, $J_1=14.6$ $J_2=2.4$ Hz), 1.66 (1H, dd, $J_1=14.6$ $J_2=10.0$ Hz), 4.21 (1H, dd, $J_1=12.3$ $J_2=10.0$ $J_3=6.2$ $J_4=2.4$ Hz)

Compound 17a Colourless oil

λ	598	578	546	436	365
α	-2.7	-3.0	-3.3	-8.0	-18.0

(CHCl_3 , c 1.02%) IR ν_{\max} cm^{-1} 3060, 1730, 1635, 1230, 1020, 930, 880, 800; ^1H NMR (CDCl_3) δ 0.73 (3H, s), 1.01 (6H, d, $J=6.9$ Hz), 2.04 (3H, s), 4.71 (1H, dd, $J_1=11.8$ $J_2=4.5$ Hz), 4.66 (1H, d, $J=1.4$ Hz), 4.85 (1H, d, $J=1.4$ Hz), 5.27 (1H, m) ^{13}C NMR (Table 1)

Compound 18 Semisolid product.

λ	589	578	546	436
α	277.0	290.7	337.2	639.8

(CHCl_3 ; c 1.01%) IR ν_{\max} cm^{-1} 3100, 2220, 2120, 1750, 1650, 1630, 1580, 1230, 1030, 970, 915, 850, 800. ^1H NMR and ^{13}C NMR, see Tables 2 and 3

REFERENCES

- Pascual Teresa, J. de, Gonzalez, M. S., De Dios, M. A., San Segundo, J. M., Vicente, S. and Bellido, I. S. (1981) *Riv. Ital.* **62**, 355
- Kirmse, W., Knist, J. and Ratajczak, H. (1976) *Chem. Ber.* **109**, 2296.
- Herz, W., Bath, S. V. and Santhanam, P. S. (1970) *Phytochemistry* **9**, 891.
- Shukla, Y. N., Sokolowski, E. A., Fales, H. M. and Kapadia, G. J. (1976) *Phytochemistry* **15**, 1788
- Cussans, M. J. and Huckerby, T. N. (1975) *Tetrahedron* **31**, 2719.
- Forgacs, P., Desconclois, J. F., Pousset, J. L. and Rabaron, A. (1978) *Tetrahedron Letters* 4783.
- Pascual Teresa, J. de, Vicente, S., Gonzalez, M. S. and Bellido, I. S. (1983) *Phytochemistry* **22**, 2235.
- Pascual Teresa, J. de, Bellido, I. S., Gonzalez, M. S. and Vicente, S. (1984) *Phytochemistry* **23**, 2064
- Pascual Teresa, J. de, Bellido, I. S., Gonzalez, M. S. and Vicente, S. (1986) *Phytochemistry* **25**, 185.
- Banarjee, S., Grenz, M., Jakupovic, J. and Bohlmann, F. (1985) *Planta Med.* **197**
- Bohlmann, F., Ates, N., King, R. M. and Robinson, H. (1983) *Phytochemistry* **22**, 1675.
- Howard, B. M. and Fenical, W. (1977) *J. Org. Chem.* **42**, 2519
- Minato, H. and Ishikawa, M. (1967) *J. Chem. Soc. (C)* 424
- Bohlmann, F., Burkhardt, T. and Zdero, C. (1973) *Naturally Occurring Acetylenes*. Academic Press, London
- Zeisberg, R. and Bohlmann, F. (1974) *Chem. Ber.* **107**, 3800.
- Martinez, V., Barbera, O., Sanchez-Parareda, J. and Alberto Marco, J. (1987) *Phytochemistry* **9**, 2619

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KANSHONES D AND E, SESQUITERPENOIDS OF *NARDOSTACHYS CHINENSIS* ROOTS*

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Key Word Index—*Nardostachys chinensis*; Valerianaceae; sesquiterpenoid; kanshones D and E.

Abstract—Two new minor sesquiterpenoids, kanshones D and E, were isolated from *Nardostachys chinensis* along with isonardosinone, nardosinone diol, and nardofuran. The structures of kanshones D and E were elucidated by spectral means together with chemical transformation.

The crude drug, 'kanshoko' (Chinese spikenard), prepared from the rhizomes and roots of the valerianaceous plant *Nardostachys chinensis* Batalin (Valerianaceae), is used in the Oriental system of medicine for stomachic and sedative purposes, and is known to elaborate a number of

sesquiterpenoids [1-11]. During the phytochemical investigation of this plant, we have so far isolated an iridoid, nardochin, and sesquiterpenoids, kanshones A, B and C, together with its known constituent nardosinone [12-14]. Further reinvestigation of the methylene chloride extract of this plant material has resulted in the isolation of two minor sesquiterpenoids kanshones D (1) and E (2) in addition to isonardosinone (3), nardosinone diol (4) and nardofuran (5), the latter two of which were previously reported as reaction products of nardosinone

* Part 63 in 'Sesquiterpenoids'. Also Part 130 in 'The Validity of the Oriental Medicines'.

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[5, 15] The advancement of the structures for kanshones D and E were mainly based on spectral data and chemical transformation

Kanshone D, was shown to possess the molecular formula $C_{15}H_{22}O_4$ on the basis of its molecular ion peak at m/z 266 and its ^{13}C NMR spectrum. The UV absorption maximum at 251 nm and IR spectral bands at 3450 and 1698 cm^{-1} inferred that kanshone D has a hydroxyl group and a conjugated carbonyl function. In the 1H NMR spectrum, signals due to hydrogens of an ABX type (δ 2.34 (1H, d , $J = 6.2$ Hz), 3.40 (1H, d , $J = 3.2$ Hz) and 3.65 (1H, dd , $J = 6.2$ and 3.2 Hz)) as well as one secondary methyl and three tertiary methyl groups appeared essentially at the same positions as those of C-6, C-8, C-7 and four methyl hydrogen signals of isonardosinone (3), pointing out a structural similarity between these two compounds. However, kanshone D exhibited an olefinic hydrogen signal at δ 6.84 as a doublet (1H, $J = 5.7$ Hz), whereas isonardosinone (3) displayed the corresponding signal at δ 6.97 as a triplet (1H, $J = 4.2$ Hz). Moreover, an extra carbinyl hydrogen signal at δ 4.22 (1H, m), which was shifted downfield to δ 5.22 (1H, m) in its monoacetate (1a), was also discernible in the 1H NMR spectrum of kanshone D. These findings, in connection with the difference in the molecular formula between kanshone D and isonardosinone (3) by an extra oxygen atom, revealed that kanshone D bears a hydroxyl group at C-2. The orientation of this hydroxyl group was shown to be spatially close to the C-4 hydrogen from the distinct downfield shift ($\Delta\delta$ 0.40 ppm) of the C-4 methine hydrogen signal at δ 2.95 (1H, m) and the significant upfield shift ($\Delta\delta = 7.6$ ppm) of the C-4 carbon signal at δ 25.5 (d) as compared to those of isonardosinone (3) (δ 2.55 (1H, m) and δ 33.1 (d), respectively). The Cotton effects in the CD spectrum of kanshone D have the identical signs to those of isonardosinone (3), inferring that both compounds have the same absolute configurations. Accordingly, kanshone D was found to be 1, which is also confirmed by comparison of ^{13}C NMR spectrum of kanshone D with that of isonardosinone (3) not reported earlier (Table 1).

Kanshone E (2), obtained as a gummy residue, was proved to have the molecular formula $C_{15}H_{20}O_4$ from its mass spectral peak at m/z 264 and an analysis of its ^{13}C NMR spectrum. Like its other congeners the 1H NMR spectrum spoke the presence of hydrogens of an ABX type (δ 3.72 (1H, dd , $J = 6.2$ and 3.3 Hz), 3.51 (1H, d , $J = 3.3$ Hz) and 2.48 (1H, d , $J = 6.2$ Hz), one secondary methyl and three tertiary methyl groups in kanshone E. Further, an olefinic hydrogen signal at δ 6.69 as a singlet

and two carbonyl carbon signals at δ 199.3 and 193.7 (each s) led to the conclusion that kanshone E is the dehydro derivative of kanshone D. In order to confirm the alleged structure as well as the absolute configurations on chiral centers of kanshone E, allylic oxidation of kanshone D was carried out to afford a compound which was identical with kanshone E (TLC, mass and 1H NMR spectra), revealing identical stereochemistry for these two compounds. The structure of kanshone E thus deduced was well corroborated with its ^{13}C NMR spectrum (Table 1).

EXPERIMENTAL

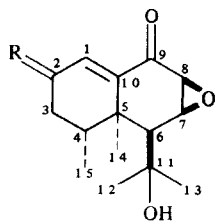
Isolation of kanshones D and E Dried rhizomes and roots (4.5 kg) of *Nardostachys chinensis* were extracted with CH_2Cl_2 (61×3) to afford a brown oil (200 g), which was chromatographed over silica gel (1 kg). The column was eluted with n -hexane and n -hexane-EtOAc mixtures of increasing polarity. Rechromatography of one of the portions of the n -hexane-EtOAc (1:1) eluates (100 g) of the above column over silica gel (0.5 kg) using hexane-Me₂CO (4:1) yielded isonardosinone, nardosinone diol, nardofuran followed by kanshone D (1) as colourless needles (12 mg) and gummy kanshone E (2) (17 mg).

Table 1 ^{13}C NMR spectra of 1-5 (250 MHz, $CDCl_3$)

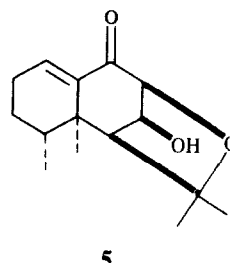
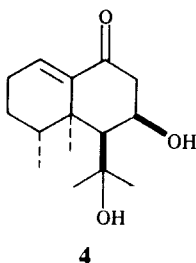
C	1	2	3	4†	5
1	134.0 d	130.0 d	139.4 d	136.7 d	137.3 d
2	61.5 d	*199.3 s	*26.0 t	*26.3 t	*25.5 t
3	33.7 t	42.1 t	*26.4 t	*25.5 t	*26.2 t
4	25.5 d	32.8 d	33.1 d	32.8 d	31.6 d
5	43.0 s	44.7 s	43.3 s	40.3 s	39.8 s
6	48.6 d	50.1 d	49.8 d	52.7 d	56.0 d
7	*52.8 d	*53.1 d	*53.4 d	68.3 d	75.7 d
8	*56.4 d	*56.8 d	*56.8 d	46.2 t	87.8 d
9	193.9 s	*193.7 s	194.0 s	200.6 s	197.7 s
10	142.4 s	155.6 s	140.6 s	142.1 s	141.8 s
11	74.7 s	75.3 s	75.1 s	77.9 s	84.9 s
12	*30.9 q	*32.7 q	*31.9 q	*37.6 q	*31.6 q
13	*28.6 q	*27.8 q	*27.7 q	*29.5 q	*31.9 q
14	21.5 q	21.8 q	24.9 q	26.0 q	24.3 q
15	16.0 q	17.4 q	17.3 q	16.2 q	15.2 q

* Data can be interchanged within the group

† 500 MHz



- 1 R = α -H, β -OH
 1a R = α -H, β -OAc
 2 R = O
 3 R = H₂



Kanshone D (1) Colourless needles, mp 152–153°, $[\alpha]_D + 57.9^\circ$ (CHCl_3 ; c 0.52), CD $[\theta]_{350} -600$, $[\theta]_{247} +2560$ (dioxane; c 0.119), EIMS (direct inlet) 70 eV, m/z 266 $[\text{M}^+]$, 234, 190, 180, 174, 161, 160, 151, 133, 120, 91, 69, 59, UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ) 251 (3.67), IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} 3450, 1698, 1635, 1470, 1268, 1070, $^1\text{H NMR}$ (100 MHz, CDCl_3) δ 0.98 (3H, s, H-14), 1.00 (3H, d, $J = 6.5$ Hz, H-15), 1.45, 1.50 (3H, each s, H-12, H-13), 2.34 (1H, d, $J = 6.2$ Hz, H-6), 2.95 (1H, m, H-4), 3.40 (1H, d, $J = 3.2$ Hz, H-8), 3.65 (1H, dd, $J = 6.2$ and 3.2 Hz, H-7), 4.22 (1H, m, H-2), 6.84 (1H, d, $J = 5.7$ Hz, H-1)

Kanshone E (2) Gummy residue, $[\alpha]_D -42.2^\circ$ (CHCl_3 , c 1.29); EIMS (direct inlet) 70 eV, m/z 264 $[\text{M}^+]$, 247, 246, 231, 228, 219, 217, 189, 162, 149, 136, 122, 120, 91, 85, 76, 71, 69, 67, 65, 59; UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ) 253 (3.68), IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} 3530, 1706, 1688, 1470, 1400, 1380, 1300, 1258, $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 1.09 (3H, d, $J = 6.0$ Hz, H-15), 1.26, 1.33, 1.52 (3H, each s, H-14, H-12, H-13), 2.25 (1H, dd, $J = 16.0$ and 14.0 Hz, H-3), 2.31 (1H, dd, $J = 16.0$ and 4.0 Hz, H-3), 2.48 (1H, d, $J = 6.2$ Hz, H-6), 3.25 (1H, m, H-4), 3.51 (1H, d, $J = 3.3$ Hz, H-8), 3.72 (1H, dd, $J = 6.2$ and 3.3 Hz, H-7), 6.69 (1H, s, H-1)

Acetylation of kanshone D A mixture of kanshone D (3 mg), Ac_2O (0.25 ml) and pyridine (0.25 ml) was kept under dry condition for overnight at room temp. Usual work-up afforded gummy kanshone D monoacetate (**1a**) (2.5 mg); EIMS (direct inlet) 70 eV m/z 306 $[\text{M}^+]$, 264 $[\text{M}-42]^+$, 248, 189, 174, 162, 161, 133, 90, 78, 76, 59, 42, $^1\text{H NMR}$ (500 MHz, CDCl_3) δ : 0.99 (3H, d, $J = 6.3$ Hz, H-15), 1.00, 1.38, 1.50 (3H, each s, H-14, H-12, H-13), 2.00 (3H, s, $-\text{OAc}$), 2.95 (1H, m, H-4), 3.40 (1H, d, $J = 4.0$ Hz, H-8), 3.65 (1H, br s, H-7), 5.22 (1H, m, H-2), 6.84 (1H, s, H-1)

Allylic oxidation of kanshone D A mixture of kanshone D (2.5 mg) and activated MnO_2 (300 mg) in Me_2CO (3 ml) was stirred for overnight at room temp. to afford kanshone E, which was identical in TLC, mass, and $^1\text{H NMR}$ spectra with natural kanshone E

REFERENCES

1. Schulte, K. E., Rucker, G. and Glauch, G. (1967) *Planta Med.* **15**, 274
2. Rucker, G. (1968) *Liebigs Ann. Chem.* **717**, 221
3. Rucker, G. (1969) *Chem. Ber.* **102**, 2691
4. Rucker, G. (1969) *Chem. Ber.* **206**, 2697
5. Rucker, G. (1970) *Chem. Ber.* **102**, 2707
6. Rucker, G. (1970) *Planta Med.* **19**, 16
7. Rucker, G. (1970) *Liebigs Ann. Chem.* **733**, 152
8. Rucker, G. and Kretzschmar, U. (1971) *Liebigs Ann. Chem.* **748**, 214
9. Hikino, H., Hikino, Y., Koakutsu, S. and Takemoto, T. (1972) *Phytochemistry* **20**, 2097
10. Shide, L., Mayer, R. and Rucker, G. (1987) *Planta Med.* **53**, 332.
11. Shide, L., Olbrich, A., Mayer, R. and Rucker, G. (1987) *Planta Med.* **53**, 556
12. Bagchi, A., Oshima, Y. and Hikino, H. (1988) *Planta Med.* **54**, 87
13. Bagchi, A., Oshima, Y. and Hikino, H. (1988) *Phytochemistry* **27**, 1199
14. Bagchi, A., Oshima, Y. and Hikino, H. (1988) *Phytochemistry* (in press)
15. Rucker, G. and Kahrs, K. H. (1973) *Liebigs Ann. Chem.* **432**